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# Signed Causal Bayesian Networks for Microbiomes

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## Abstract

1        Inferring causality is the process of connecting causes with effects. Identifying  
2        even a single causal relationship from data is more valuable than observing dozens  
3        of correlations in a data set. Microbe-microbe and host-microbe interactions play a  
4        vital role in both health and disease. In this study, we investigate how to learn a  
5        causal structure from data from microbiome studies and its potential interpretation  
6        about events and processes in the microbial community under study. We report  
7        evidence that causal structure can extract colonization patterns even though the  
8        analysis only uses data with no temporal information.

## 9    1 Introduction and Motivation

10    Causation is an important type of relationship to be explored with biological data. Thus, it makes  
11    sense to see if causal Bayesian networks can identify relationships that are suggestive of causation,  
12    leading to lab experiments for validation. Bayesian networks (BNs) were used by Zhang et al.  
13    to understand changes in gene regulatory networks (1), and Szal et al. used BNs to understand  
14    relationships among taxa in microbiomes (2). By modeling metabolic reactions and their involvement  
15    in multiple subnetworks of “metabosystems”, Shafiei et al. used BNs to infer differential prevalence  
16    of metabolic subnetworks within microbial communities (3).

17    A *microbiome* is a community of microbes including bacteria, archaea, protists, fungi and viruses that  
18    share an environmental niche (4). Microbiomes can be modeled as a *social network* because of the  
19    complex set of potential interactions between its various taxonomic members (5; 6). To understand  
20    potential interactions between taxa in a microbial community, the construction of co-occurrence  
21    networks (CoN) was proposed by Fernandez et al. (5) and Faust et al. (7). The results suggested that  
22    the reason groups of taxa frequently co-infect cohorts of subjects or did the opposite, i.e., co-avoided  
23    cohorts of subjects, was because of underlying interactions between them. Unfortunately, that is as  
24    far as CoNs are able to go in terms of inferring complex relationships in microbiomes.

25    In this paper, we investigate how to infer directional relationships between microbial taxa in a  
26    microbiome. In humans, normal microbial colonization starts from birth and with the passage of time  
27    these communities become relatively stable (8). Some microbes recruit others suggesting an order of  
28    colonization in many microbial communities. Understanding colonization and its order can provide  
29    a window into how infections take hold. We show that causal structure or *signed Causal Bayesian*  
30    *Networks* (scBNs), a variant of BNs obtained by combining BNs with Co-occurrence networks  
31    can help tease apart some of these directed relationships and provide a glimpse into the complex  
32    and dynamic world of microbial communities. Work is underway to investigate how to infer other  
33    causal relationships from the same data. In particular, our work will highlight the microbial players  
34    involved in recruiting other microbes, the key players in causing disease, their relative importance  
35    in the disease process, the role of beneficial microbes in alleviating disease symptoms, the role of  
36    metabolites in disease, the identification of potential targets for treatment, and much more.

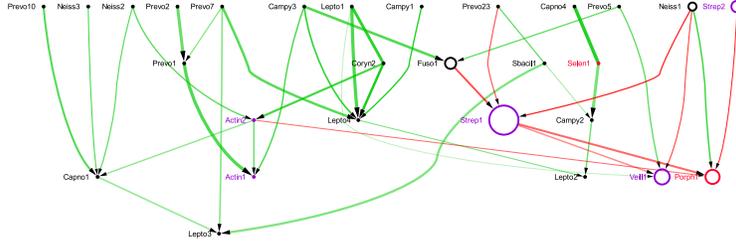


Figure 1: **Signed Causal Bayesian Network**. Nodes represent microbial taxa, edges represent the relationships among taxa, red and green edges represent negative and positive correlation respectively.

## 37 2 Methods

38 *Causal structures* (CS) are a class of *Probabilistic Graphical Models* (PGMs) (9; 10) where each  
 39 node represents a random variable from a set,  $\mathbf{X} = \{X_i, i = 1, \dots, n\}$  with  $n$  random variables.  
 40 These structures are represented as a graph  $G = (V, E)$ , where each vertex in  $V$  represents a random  
 41 variable from  $\mathbf{X}$ , and  $E$  is the set of edges on  $V$ . The graph  $G$  is also known as a *causal Bayesian*  
 42 *network* on  $X$ . Although undirected edges are used in cases where the direction cannot be reliably  
 43 determined or when both directions appear to be valid, the graph  $G$  is often “manipulated” to be a  
 44 Directed Acyclic Graph (DAG). Each random variable  $X_i$  has an associated probability distribution. A  
 45 directed edge in  $E$  between two vertices represents direct stochastic dependencies. Therefore, if there  
 46 is no edge connecting two vertices, the corresponding variables are either marginally independent or  
 47 conditionally independent (conditional on the rest of the variables, or some subset thereof). To learn  
 48 a causal structure we adopted a conditional independence test based method proposed by Spirtes  
 49 et al. (11), later modified by Colombo and Maathuis to make it *order independent*, and known as  
 50 *PC-Stable* algorithm (12). PC-stable consists of three steps - adjacency search in order to learn the  
 51 “skeleton”, identifying important substructures called *v-structures*, and detecting and orienting other  
 52 arcs. In Step 1, the algorithm starts with a complete undirected graph and then performs a series of  
 53 conditional independence tests to eliminate as many edges as possible. The remaining undirected  
 54 graph is referred to as the *skeleton*. Step 2 is key to inferring a directional model, and uses the concept  
 55 of *v-structures* (13). Step 3, three rules (12) are applied repeatedly to orient remaining undirected  
 56 edges (i.e., arcs not in *v-structures*). Finally, we added sign information from CoNs to the edges in  
 57 the causal structure.

58 We used oral data sets (16S rRNA sequences) generated as part of the Human Microbiome Project  
 59 (HMP) from eight different sites within the oral cavity from 242 healthy adults (129 males, 113  
 60 females) (14; 4). The samples included: saliva, buccal mucosa (cheek), keratinized gingiva (gums),  
 61 palatine tonsils, throat, tongue dorsum, and supra- and sub-gingiva dental plaque (tooth biofilm above  
 62 and below the gum). Abundance of individual taxa were computed after amplification of a specific  
 63 hypervariable region of the bacterial 16S rRNA gene, followed by sequencing, grouping reads into  
 64 common Operational Taxonomic Units (OTUs) and quantification (15). Mothur (16) was used to  
 65 compute the microbial abundance profile.

## 66 3 Results and Discussion

67 Figure 1 shows a signed causal structure learned from keratinized gingiva data set. The results  
 68 with all oral data sets showed a surprising connection to the order in which microbes colonize the  
 69 human mouth. Two significant observations were as follows. (1) The directed edges of scBN for  
 70 the oral microbiome data set were consistent with the colonization order. A total of 716 edges were  
 71 generated for the oral microbiome scBN with the colonization order known for 78 edges. Only 2  
 72 edges in the scBN were inconsistent with the known direction, Resulting in an accuracy of 97.4%.  
 73 (2) scBN Edges with negative correlations were consistent with the colonization order (early to late  
 74 colonizers). All directed edges between two taxa from two colonization groups were negatively  
 75 correlated. Thus, the scBNs help us to infer potential relationships and dependencies within a  
 76 microbiome, and the colonization order without time information. scBNs could help in understanding  
 77 the other dependencies among the entities of a microbial community.

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